

Nucleotide Sequence Tankyrase homologue isotype1

CTTTGAAGACACTGGATTTTCATACTTTTGCCTGGGGTTATCTCTCTGTGTCTCACTACATAGACAAATA
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 AA

FIGURE 2: SEQ ID NO:2

Nucleotide Sequence Tankyrase homologue isotype2

CGCGCTGCTCCGCCCCGCCGCGGGGCGAGCCGGGGGGCAGGGAGCCCAGCGAGGGGCGCGCTGGGCGCGG
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 CCGAGGCCGTGGAGCCGGCCGCCGAGAGCTGTTGAGGCGTGCCGCAACGGGGACGTGGAACGAGTCA
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TAGTTATTTTAAGATTAATTCCACTGAACCTAAAATCATCAAAGCAGCGGCCTCTACGTTTTTAC
TCCTTTGCTGAAAAA

FIGURE 3: SEQ ID NO:3

Amino Acid Sequence Tankyrase homologue isotype1

GFGRKDVVEYLLQNGASVQARDDGGLIPLHNACSFGHADEVNLLLRHGADPNARDNWNYPPLHEAAIKG
KIDVCIVLLQHGAEP TIRNTDGR TALDLADPSAKAVLTGEYKKDELLESARSGNEEKMMALLTPLNVNC
HASDGRKSTPLHLAAGYNRVKIVQ LLLQH GADVHAKDKGDLVPLHNACSYGHYEVTELLVKHGACVNAM
DLWQFTPLHEAASKNRVEVCSLLLSYGADPTLLNCHNKS AIDLAPTPQLKERLAYEFKGHSLQAAREA
DVTRIKKHL SLEMVNF KHPQTHETALHCAAASPYPKRKQICELL LRKGANINEKTKEFLTPLHVASEKA
HNDVVEVVVKHEAKVNALDNLGQTS LHRAAYCGHLQTCRLLLSYGCDPNIISLQGF TALQMGNENVQQL
LQEGISLGNSEADRQLLEAAKAGDVETVKKLCTVQSVNCRDIEGRQSTPLHFAAGYNRVSVVEYLLQHG
ADVHAKDKGGLVPLHNACSYGHYEAELLVKHGAVVNVADLWKFTPLHEAAAKGKYEICKLLLQH GADP
TKKNRDGNTPLDLVKDGD TDIQDLLRGDAALLDAAKKGCLARVKKLSSPDNVNCRDTQGRHSTPLHLA
GYNNLEVAEYLLQH GADVNAQDKGGLIPLHNAASYGHVDVAALLIKYNACVNATDKWAFTPLHEAAQKG
RTQLCALLLAHGADPTLKNQEGQTPDLV SADDVSALLTAAMPSPALPSCYKPQVLNGVRSPGATADAL
SSGPSSPSSLSAASSLDNLSGSFSELSSVSSSGTEGASSLEKKEVPGVDFSITQFVRNLGLEHLMDIF
EREQITLDV LVEMGHKELKEIGINAYGHRHKL IKGVERLISGQQGLNPYLTNTSGSGTILIDLSPDDK
EFQSV EEE MQSTVREHRDGGHAGGIFNRYN ILKIQKVCNKKLWERYTHRRKEVSEENHNHANERMLFHG
SPFVNAI IHKGFDERHAYIGGMFGAGIYFAENSSSKSNQYVYGIGGGTGCPVHKDRSCYICHRQLLFCRV
TLGKSFLQFSAMKMAHSPPGHHSVTGRPSV

Bold = potential starting methionine

FIGURE 4: SEQ ID NO:4

Amino Acid Sequence Tankyrase homologue isotype2

RCSARRGAAGGQGAQRGARVGA AHGTAPDPVTAGSQAARALSASSPGGLALLLAGPGLLLRLLALLLAV
AAARIMSGRR CAGGGAACASAAAEAVEPAARELFEACRNGDVERVKRLVTPEKVNSRD TAGRKSTPLHF
AAGFGRKDVVEYLLQNGANVQARDDGGLIPLHNACSFGHADEVNLLLRHGADPNARDNWNYPPLHEAAI
KGKIDVCIVLLQHGAEP TIRNTDGR TALDLADPSAKAVLTGEYKKDELLESARSGNEEKMMALLTPLNV
NCHASDGRKSTPLHLAAGYNRVKIVQ LLLQH GADVHAKDKGDLVPLHNACSYGHYEVTELLVKHGACVN
AMDWQFTPLHEAASKNRVEVCSLLLSYGADPTLLNCHNKS AIDLAPTPQLKERLAYEFKGHSLQAAR
EADVTRIKKHL SLEMVNF KHPQTHETALHCAAASPYPKRKQICELL LRKGANINEKTKEFLTPLHVASE
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QLLQEGISLGNSEADRQLLEAAKAGDVETVKKLCTVQSVNCRDIEGRQSTPLHFAAGYNRVSVVEYLLQ
HGADVHAKDKGGLVPLHNACSYGHYEAELLVKHGAVVNVADLWKFTPLHEAAAKGKYEICKLLLQHGA
DPTKKNRD GNTPLDLVKDGD TDIQDLLRGDAALLDAAKKGCLARVKKLSSPDNVNCRDTQGRHSTPLHL
AAGYNNLEVAEYLLQH GADVNAQDKGGLIPLHNAASYGHVDVAALLIKYNACVNATDKWAFTPLHEAAQ
KGRTQLCALLLAHGADPTLKNQEGQTPDLV SADDVSALLTAAMPSPALPSCYKPQVLNGVRSPGATAD
ALSSGPSSPSSLSAASSLDNLSGSFSELSSVSSSGTEGASSLEKKEVPGVDFSITQFVRNLGLEHLMD
IFEREQITLDV LVEMGHKELKEIGINAYGHRHKL IKGVERLISGQQGLNPYLTNTSGSGTILIDLSPD
DKEFQSV EEE MQSTVREHRDGGHAGGIFNRYN ILKIQKVCNKKLWERYTHRRKEVSEENHNHANERMLF
HGSPFVNAI IHKGFDERHAYIGGMFGAGIYFAENSSSKSNQYVYGIGGGTGCPVHKDRSCYICHRQLLFC
RVTLGKSFLQFSAMKMAHSPPGHHSVTGRPSV NGLALAEYVIYRGEQAYPEYLITYQIMRPEGMVDG

FIGURE 5

Schematic Presentation of Dominant negative Mutants for Tankyrase Homologue

Dominant Negative Mutants:

Truncation: 429 Δ C- of the C-terminal catalytic domain – truncation of the catalytic domain of PARP acts as a dominant negative when overexpressed *in vivo* (Oncogene 1999 Nov 25;18(50):7010-5)
 Point mutant: E945A Δ C- conserved residue in PARP domain, thought to be important in NAD⁺ binding

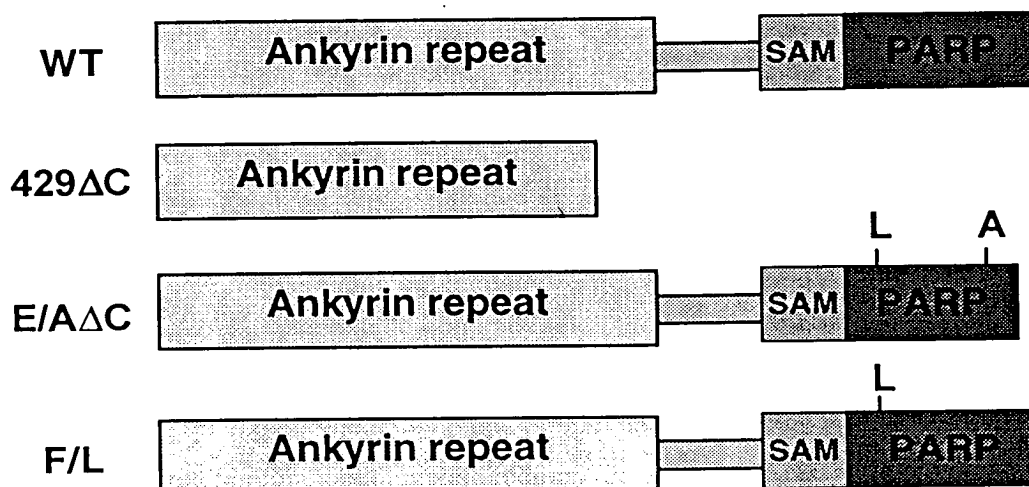
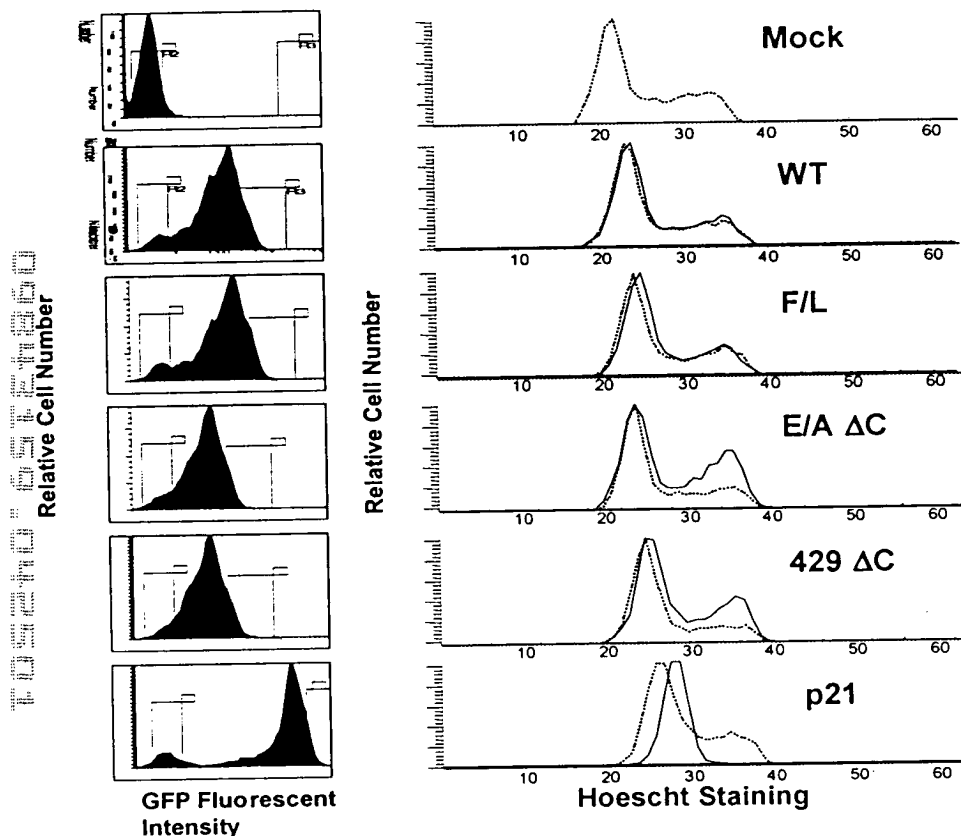


FIGURE 6

Cell Cycle Analysis of A549 Cells Infected With GFP-fused Wild Type and Mutant Tankyrase Homologue



Gates for GFP positive cells and negative cells are shown in FACS analysis of GFP expression

Cells were infected retrovirus encoding GFP-fused wild type and mutant Tankyrase homologue and incubated for 48 hours.

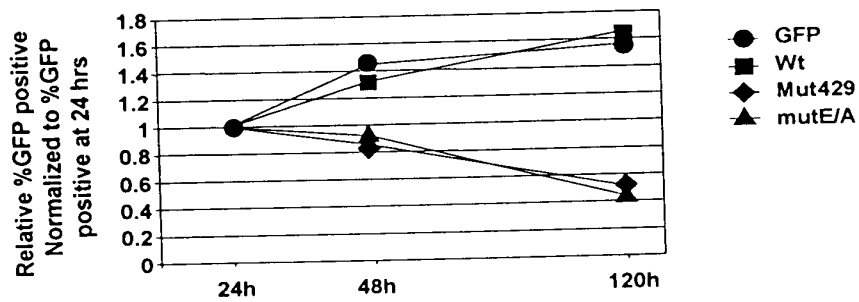
— Cell cycle analysis of GFP positive cells

..... Cell cycle analysis of GFP negative cells

FIGURE 7

Kinetics of GFP Positive cells in A549 Cells and Human Mammary Epithelial Cells(HMEC) After Retrovirus Infection Encoding GFP-fused Wild Type and Mutant Tankyrase Homologue

A549



HMEC

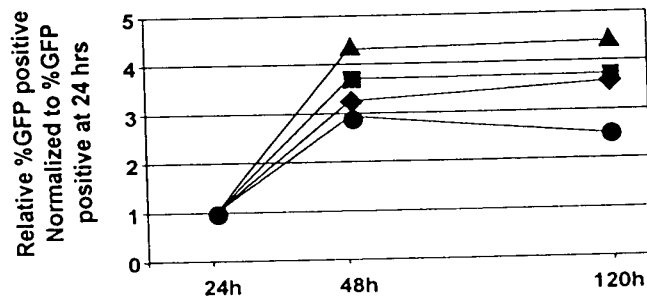
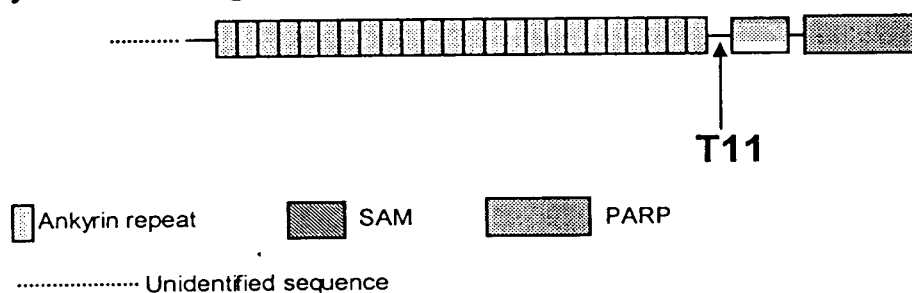


FIGURE 8

The Binding Site of Antisense Oligos Against Tankyrase Homologue

Tankyrase Homologue



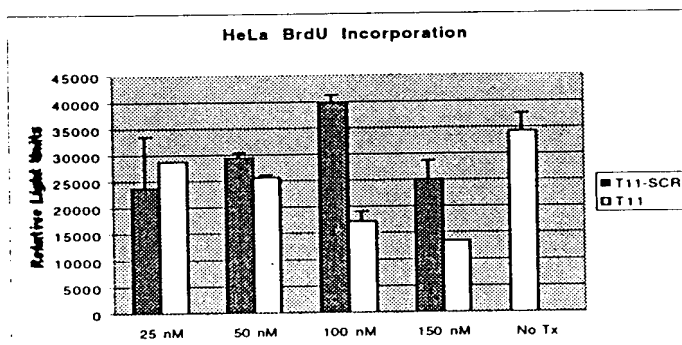
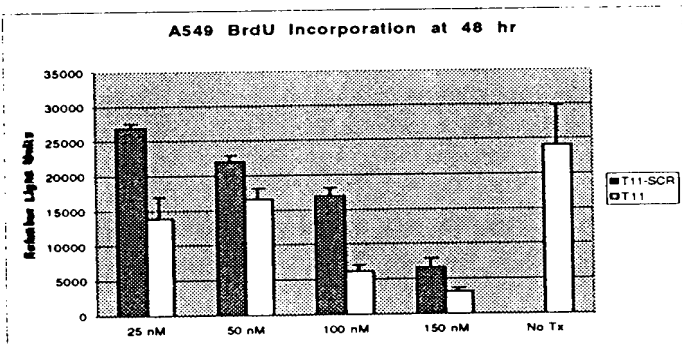
T11

	GTGGAACAGAGGGTGCTTCC	
Tankyrase Homologue	GTGGAA ^T CAGAGGGTGCTTCCAGTTTGGAGAAAAAGGAGGTTCCAGGAGTAGATTTAGCAT	2838
Tankyrase	ATGCAGGGGATGGCGCCGCGGGAACAGAAAGGAAGGAAGGAGAAGTTGCTGGTCTTGACAT	3091
	* * * * *	

FIGURE 9

Anti-Proliferative Phenotype of Antisense Oligonucleotides Against Tankyrase Homologue in A549 and HeLa Cells

Proliferation Analysis



mRNA Analysis

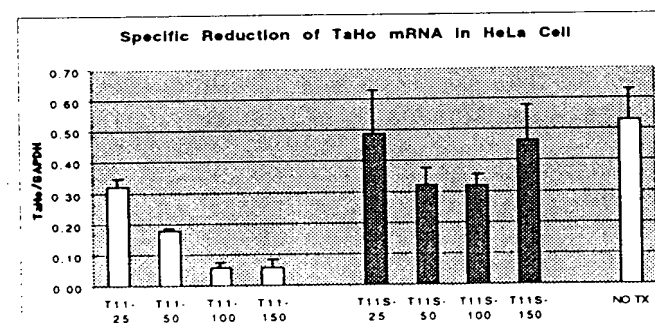
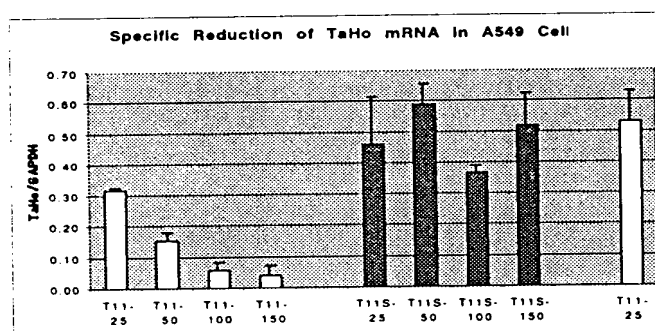
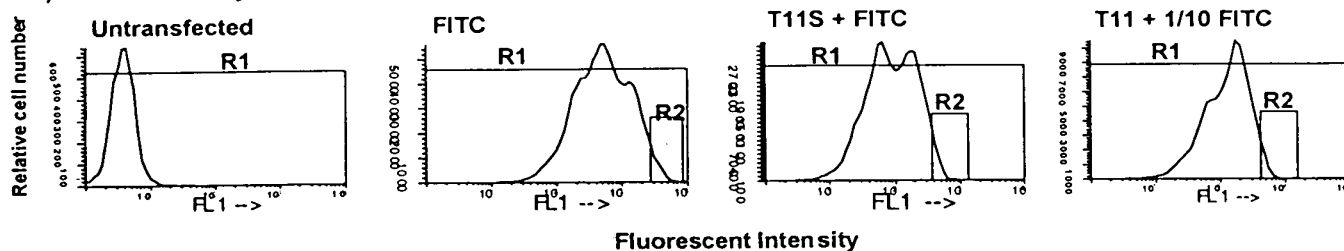


FIGURE 10

Cell Cycle Analysis of A549 Cells Transfected with Antisense Oligonucleotides Against Tankyrase Homologue at 48 Hours, Antisense Oligonucleotides (T11) and control oligonucleotides (T11S) were transfected with FITC-labeled random 20mer oligonucleotides(FITC), After 48 hours, entire population(R1) and top 5 % (R2)of FITC transfected cells were analyzed for cell cycle.

A) Gates for cell cycle analysis



B) Cell cycle analysis

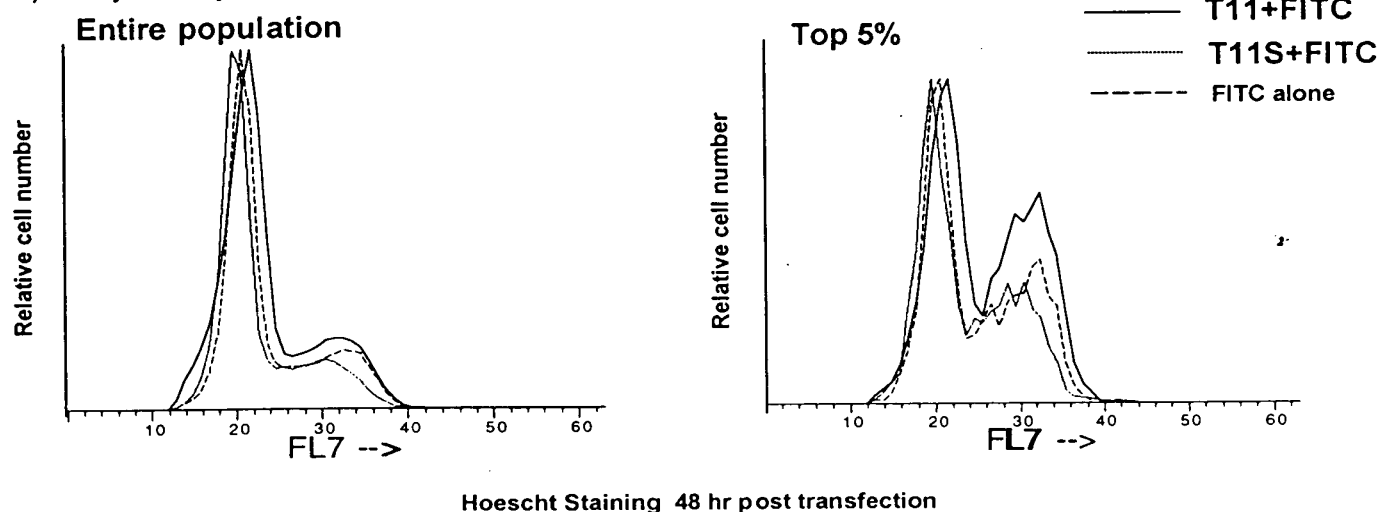


FIGURE 11

mRNA expression of Tankyrase Homologue in Several Tumors and Normal Tissues by a Taqman Analysis, mRNA expression was normalized by 90kDa Highly Basic Protein (HBP) and ribosomal protein S9 (S9).

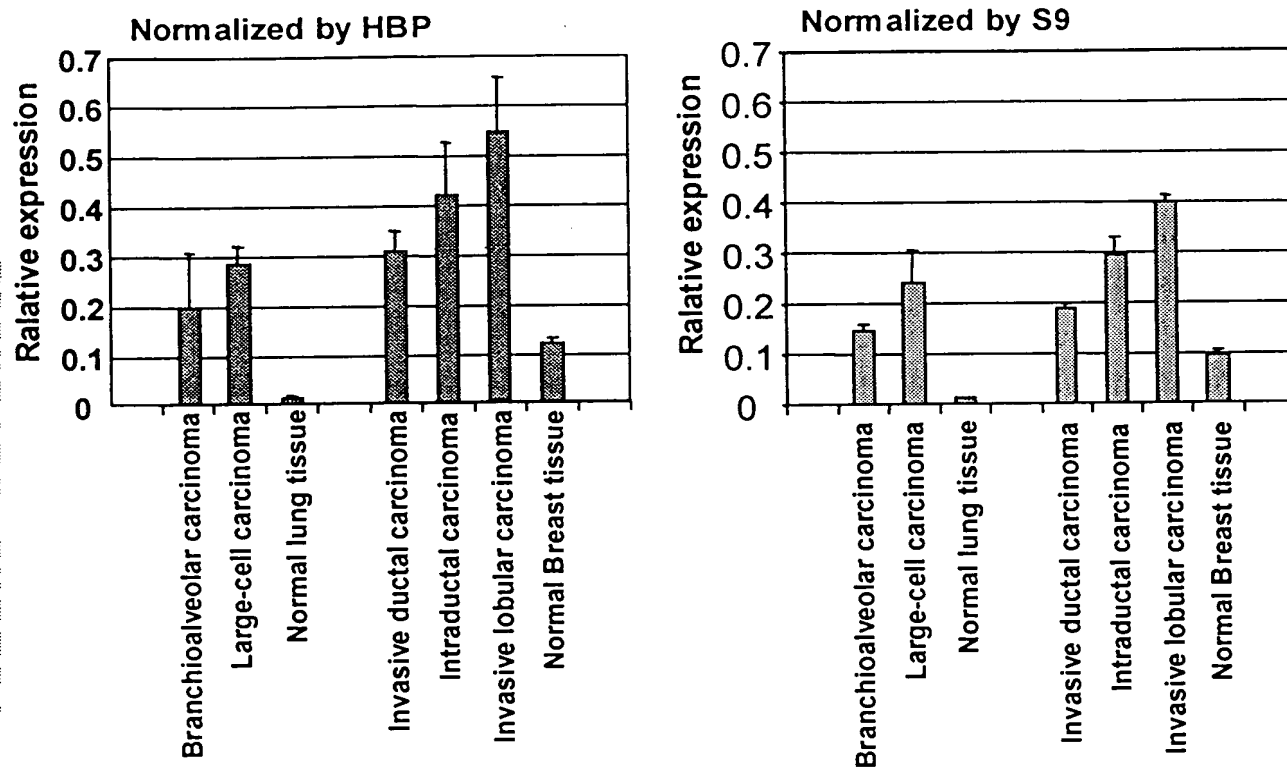


FIGURE 12

**Procedure for Nonisotopic Detection of Poly-ADP Ribosylation
Using Anti-GFP mAb-Coated Plates**

Protein lysates from 293T cells normalized by GFP
fluorescence and total protein

↓
Immobilization of GFP-tankyrase homologue in anti-
GFP Coated plates

↓
Auto PARP reaction with Biotinylated-NAD in 96 wells

↓
Detection of poly ADP ribose chains with Streptavidin-
HRP and chemiluminescent substrate

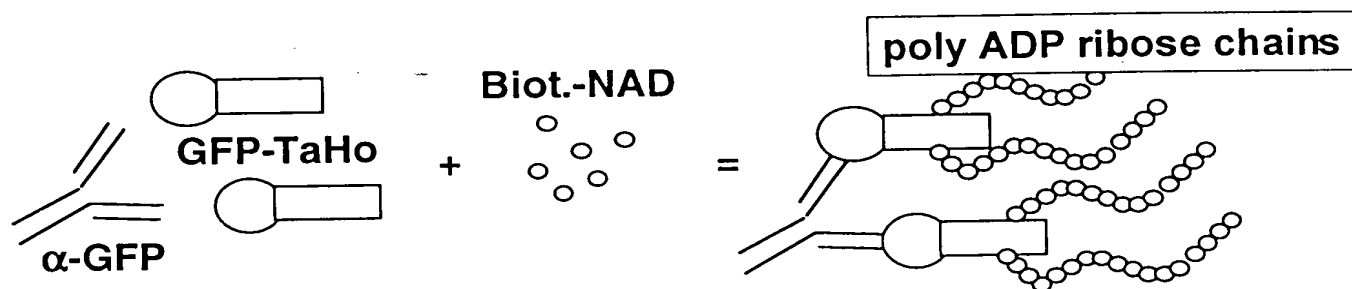


FIGURE 13

Non-Isotopic Plate-Based Detection of Taho PARP Activity in the Presence of Biotinylated NAD.

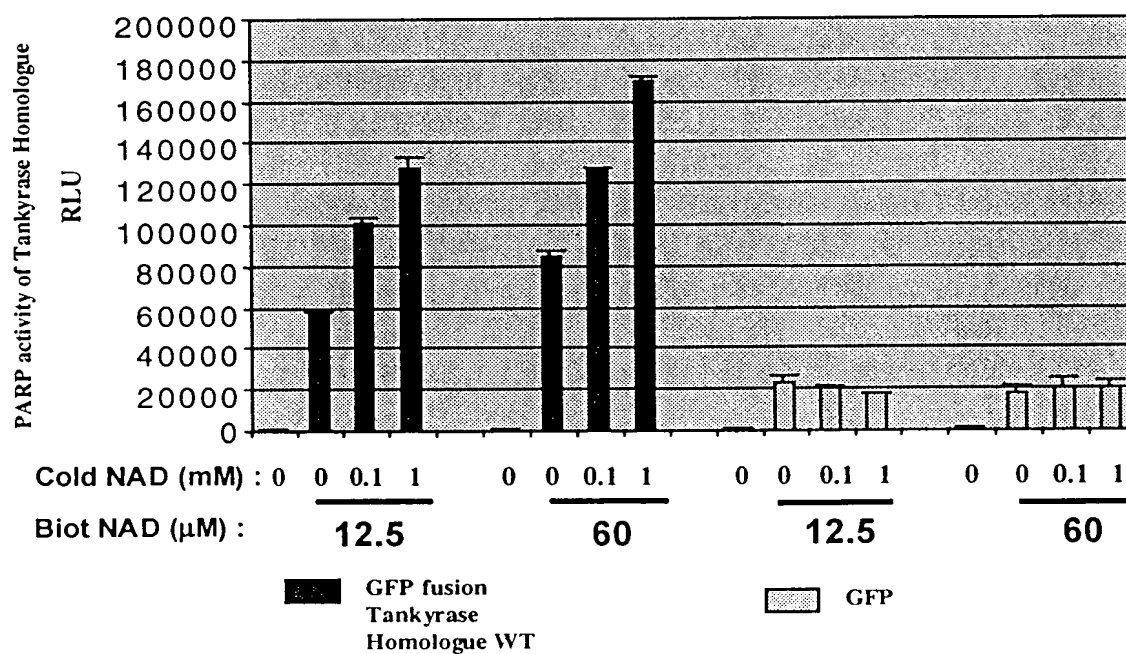


FIGURE 14

Comparison of IC₅₀ Values of the PARP Inhibitors

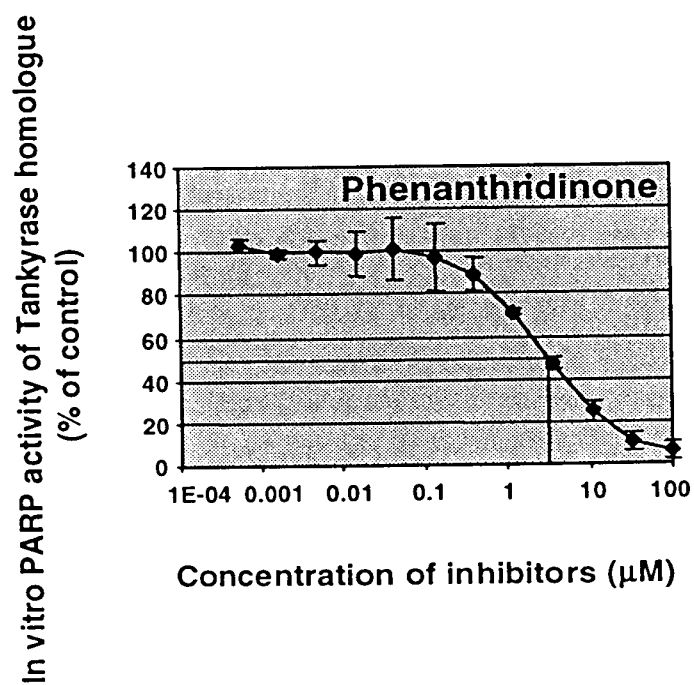
	Approximate IC ₅₀ (nM)	hPARP assay IC ₅₀ (nM)		
	<u>TaHo</u>	<u>Rigel</u>	<u>Decker *</u>	<u>Rankin *</u>
3AB	> 50 000	5 000	2 000	5 400
6(5H)Phenanthridinone	1 000-2 000	300		
Niacinamide	> 50 000	30 000	>>5 000	31 000

* Decker P et al., *Clinical Cancer Research*. 1999 May; 5:1169-1172.

* Rankin PW et al., *J Biol Chem*. 1989 Mar 15;264(8):4312-4317.

FIGURE 15

Inhibition of Tankyrase homologue PARP activity by hPARP inhibitors



TH-1: Tankyrase homologue isoform-1, TH-2: Tankyrase homologue isoform-2
M (Red): the first methionine in the sequence, Z: stop codon
In this figure, the first methionine in TH-1 sequence is position 1 (M1)

Taho C terminus deletion mutant ends at position 429 (K) and adds 28 amino acids because of frame shift.

Taho F/L mutant has the mutation at position 871

Taho E/A dC mutant has the mutation at position 948, ends at position 957 (A) and adds 2 amino acids.

TH-1	-----	
TH-2	RCSARRGAAGGQGAQRGARVGAHGTAPDPVTAGSQ	-231
TH-1	-----	
TH-2	AARALSASSPGGLALLLAGPGLLLRLLALLAVAAARIMSGRRRCAGGGAACASAAAEAVE	-171
TH-1	-----	
TH-2	PAARELFEACRNGDVERVKRLVTPEKVNSRDTAGRKSTPLHFAAGFGRKDVVEYLLQNGA	-111
	Ankyrin repeat	Ankyrin repeat
TH-1	SVQARDGGLIPLHNACSFHAEVNVNLLLRHGADPNARDNWNNTPLHEAAIKGKIDVCIV	-51
TH-2	NVQARDGGLIPLHNACSFHAEVNVNLLLRHGADPNARDNWNNTPLHEAAIKGKIDVCIV	-51
	Ankyrin repeat	Ankyrin repeat
		• TH1 start
TH-1	LLQHGAETIRNTDGR TALDPSAKAVLTGEYKKDELLESARSGNEEKMALLTPLNV	10
TH-2	LLQHGAETIRNTDGR TALDPSAKAVLTGEYKKDELLESARSGNEEKMALLTPLNV	10

FIGURE 16
(sheet 1 of 3)

	Ankyrin repeat	Ankyrin repeat
TH-1	NCHASDGRKSTPLHLAAGYNRVKIVQLLQHGADVHAKDKGDLVPLHNACSYGHYEVTEL 70	
TH-2	NCHASDGRKSTPLHLAAGYNRVKIVQLLQHGADVHAKDKGDLVPLHNACSYGHYEVTEL 70	Ankyrin repeat
TH-1	LVKHGACVNAMDLWQFTPLHEAASKNRVEVCSLLLSYGADPTLLNCHNKSAIDLAPTQL 130	
TH-2	IV	Ankyrin repeat
TH-1	KERLAYEFKGHSLLOAAREADVTRIKKHLSEMVNFKHPQTHETALHCAAAASPYPKRKQI 190	Ankyrin repeat
TH-1	CELLLRKGANINEKTKEFLTPLHVASEKAHNDVVEVVVKHEAKVNALDNLGQTSLHRAAY 250	Ankyrin repeat
TH-1	CGHLQTCRLLLSYGCDPNIISLQGF TALQMGNENVQQLLEGISLGNSEADRQLLEAACA 310	Ankyrin repeat
TH-1	GDVETVKKLCTVQSVNCRDIEGRQSTPLHFAAGYNRVSVVEYLLQHGADVHAKDKGGLVP 370	Ankyrin repeat
TH-1	LHNACSYGHYEAELLVKHGAVVNVADLWKFTPLHEAAAKGYEICKLLQLQHGADPTKKN 430	Ankyrin repeat
TH-1	GMEILLWILLKMEIQIFKICLGEMQLCZ	Ankyrin repeat
TH-1	RDGNTPLDLVKDGDTDIQDLLRGDAALLDAAKKGCLARVKKLSSPDNVNCRDQTQGRHSTP 490	Ankyrin repeat

TH-1	LHLAAGYNNLEVAEYLLQHGADVNAQDKGGLIPLHNAASYGHVDVAALLIKYNACVNATD 550 Ankyrin repeat Ankyrin repeat
TH-1	KWAFPLHEAAQKGRQTQCALLLAHGADPTLKNQEGQTPLDLSADDDVSALLTAAMPESA 610 Ankyrin repeat Ankyrin repeat
TH-1	LPSCYKPQVLNGVRSPGATADALSSGPSSPSSLSAASSLDNLGSELSVSSSGTEG 670 Ankyrin repeat
TH-1	ASSLEKKE--VPGVDFSITQFVRNLGLEHIMDIFEREQITLDVLVEMGHKELKEIGINAY 730 SAM domain
TH-1	GHRHKLIGVERLISGQQGLNPYLTLNTSGSGTILIDLSRDDKEFQSVVEEMQSTVREHR 790
TH-1	DGGHAGGIFNRYNILKIQKVCNKKLWERYTHRRKEVSEENHNHANERMLFHGSPFVNAIL 850
TH-1	HKGDERHAYIGMFGAGIYFAENSSKSNQYVYIGGGTGCPVHKDRSCYICHRQLLFCR 910 • F→L mutation PARP domain
TH-1	VTLGKSLQFSAMKMAHSPPGHHSVTGRPSVNGLALAEYVIYRGEQAYPEYLITYQIMRP 970 -----A-----LSZ •E→A •Deletion.
TH-1	EGMVVDG 976